

Curriculum Vitae
Kathryn D. Mouzakis

Department of Chemistry and Biochemistry
Loyola Marymount University
Los Angeles, CA 90045
310-338-2943
Kathryn.Mouzakis@lmu.edu

EDUCATION AND TRAINING

Postdoctoral Fellow, University of Wisconsin – Madison 05/2013 – 07/2013
Mentor: Dr. Samuel E. Butcher
Research: Structural studies of the Israeli acute paralysis virus intergenic IRES pseudoknot I domain by NMR and SAXS.

Ph.D., Biochemistry, University of Wisconsin – Madison 09/2007 – 05/2013
Research Advisor: Dr. Samuel E. Butcher
Research: Investigating HIV-1 frameshift site stem-loop structure and function.

B.S., Chemistry – Biology, Harvey Mudd College 09/2003 – 05/2007
Research Advisor: Dr. Karl A. Haushalter
Research: DNA repair efficiency, thermal stability, and substrate specificity of human 8-oxoguanine DNA glycosylase 1 variants.

ACADEMIC EMPLOYMENT

Associate Professor 08/2021 – present
Loyola Marymount University
Department of Chemistry and Biochemistry

Faculty Fellow 01/2023 – 05/2024
Loyola Marymount University
Office of Research and Creative Arts and Howard Hughes Medical Institute (HHMI) Driving Change Award

Visiting Associate Professor 06/2022 – 08/2022, 07/2023
Duke University
Department of Chemistry
Dr. Amanda Hargrove's Lab

Assistant Professor 08/2018 – 08/2021
Loyola Marymount University
Department of Chemistry and Biochemistry

Assistant Professor 08/2013 – 08/2018
Fort Lewis College
Department of Chemistry and Biochemistry

Visiting Scientist (NIH SCORE supported) 05/2017 – 07/2017, 05/2018 – 07/2018
University of Colorado School of Medicine
Department of Biochemistry and Molecular Genetics
Dr. Jeffrey S. Kieft's Lab

TEACHING/ADVISING

Courses taught: Academic Year (AY), Fall Semester (FA), Spring Semester (SP), Summer (SU):

Loyola Marymount University

Biochemistry (CHEM 370)	FA 2018, FA 2019, FA 2025, SU I 2026
Introduction to Virology (CHEM 478)	FA 2019, FA 2021, FA 2023, SP 2025, SP 2026
Directed Research (CHEM 397, CHEM 497)	SP 2023 – SP 2024, SP 2025 – SP 2026
	FA 2018 – SP 2021
Advanced Biochemistry Lab (CHEM 373)	SP 2020, SP 2021, SP 2023, SP 2024, SP 2025
Biochemistry Lab (CHEM 371)	FA 2018, FA 2020, FA 2021, FA 2023, FA 2025
Chemistry Teaching: Biochemistry (CHEM 495)	FA 2023, SP 2024, SP 2025, FA 2025
	FA 2020, SP 2021, SP 2023
Honors Thesis Advisor (HNRS 5000, BIO 598)	SP 2020, FA 2021, SP 2023, FA 2025
Chemistry Seminar (491/490/391/390)	FA 2021, FA 2025
General Chemistry II Lecture (CHEM 112)	SP 2023, SP 2024
General Chemistry II Lab (CHEM 113)	SP 2020, SU II 2020, SP 2021
Advanced Biochemistry (CHEM 372)	SP 2019, SP 2020
General Chemistry I Lab (CHEM 111)	FA 2019

Fort Lewis College

General Biochemistry II (CHEM 313)	SP 2015, SP 2017, SP 2018
Fundamentals of Chemistry I Lab (CHEM 150L)	AY 2014 – 2015, FA 2016, SP 2018
Independent Study (CHEM 299 or 499)	AY 2016 – 2017, FA 2017, SP 2018
Seminar I (CHEM 496)	FA 2017
Fundamentals of Chemistry I (CHEM 150)	AY 2013 – 2014, FA 2015, SP 2017
Special Topics in Biochemistry (CHEM 454)	FA 2016
General Biochemistry I (CHEM 311)	AY 2014 – 2015, SP 2016, FA 2016
Advanced Biochemistry Laboratory (CHEM 411)	SP 2014, SP 2016
Fundamentals of Chemistry II (CHEM 151)	SP 2016
Fundamentals of Chemistry II Lab (CHEM 151L)	FA 2015
General Biochemistry Lab (CHEM 312)	FA 2013, FA 2015
Introduction to Chemical Research (CHEM 381)	AY 2013 – 2014, AY 2014 – 2015

Advising:Loyola Marymount University

- Mentored 23 undergraduate research students.
- Mentored 84 undergraduate students enrolled in course-based undergraduate research experiences (CUREs) I developed and taught.
- Supervised 43 undergraduate research student oral/poster presentations.
- Served as an academic advisor to 3-10 Chemistry and Biochemistry majors per year, FA 2018 – present (excluding CY 2022 sabbatical and FA 2024 maternity leave).
- Served as a first-year academic advisor to 25 Chemistry and Biochemistry majors, SU 2026 – present.
- Served as an Honors Thesis advisor to 6 undergraduate research students.

Fort Lewis College

- Mentored 22 undergraduate research students.
- Mentored 31 undergraduate students enrolled CUREs I developed and taught.
- Supervised 21 undergraduate research student oral/poster presentations.

- Served as an academic advisor to 8-19 Chemistry and Biochemistry majors per year, 2013 – 2017.

SCHOLARSHIP/CREATIVE WORKS

Peer-reviewed Publications (undergraduate mentees are underlined, Corresponding Author*):

Published when Dr. Mouzakis held the rank of Associate Professor:

1. Swanson Hay, E.G.; Maille, M.M.; Zafferani, M.; Puray-Chavez, M.; Xia, M.; Luu, T.B.; Muralidharan, D.; Li, M.L.; Brewer, G.; Kutluay, S.B.; **Mouzakis, K.D.***; Hargrove, A.E.* SARS-CoV-2 Antivirals Identified from Small Molecule Modulators of Programmed –1 Ribosomal Frameshifting. *ACS Infectious Diseases* **2026**, 5(12), 1586-1599. DOI: [10.1021/acsinfecdis.5c00774](https://doi.org/10.1021/acsinfecdis.5c00774)
2. Mitchell, S.F.; **Mouzakis, K.D.*** An Approach to Transitioning Undergraduate Biochemistry Laboratory Courses Online. *The Biophysicist* **2021**, 2(2), 33-37. DOI: [10.35459/tbp.2021.000184](https://doi.org/10.35459/tbp.2021.000184)
 - Notes: this manuscript was **not included** in my application for promotion to Associate Professor submitted to the LMU Department of Chemistry and Biochemistry. It was published on September 29, 2021, after I was officially promoted on August 15, 2021.

Published when Dr. Mouzakis held the rank of Assistant Professor:

3. Thulson, E.; Hartwick, E.W.; Cooper-Sansone, A.; Williams, M.A.C.; Soliman, M.E., Robinson, L.K.; Kieft, J.S.; **Mouzakis, K.D.*** An RNA Pseudoknot Stimulates HTLV-1 *pro-pol* Programmed -1 Ribosomal Frameshifting. *RNA* **2020**, 26(4), 512-528. DOI: [10.1261/rna.070490.119](https://doi.org/10.1261/rna.070490.119)
4. **Mouzakis, K.D.**; Wu, T.; Haushalter, K.A.* Thermostability and Excision Activity of Polymorphic Forms of hOGG1. *BMC Res Notes* **2019**, 12(1), 92. DOI: [10.1186/s13104-019-4111-9](https://doi.org/10.1186/s13104-019-4111-9)
5. Au, H.H.; Cornilescu, G.; **Mouzakis, K.D.**; Burke, J.E.; Ren, Q.; Lee, S.; Butcher, S.E.; Jan, E.* Global shape mimicry of tRNA within a viral internal ribosome entry site mediates translational reading frame selection. *PNAS* **2015**, 112(47), E6446-6455. DOI: [10.1073/pnas.1512088112](https://doi.org/10.1073/pnas.1512088112)
6. **Mouzakis, K.D.**; Dethoff, E.A.; Tonelli, M.; Al-Hashimi, H.M.; Butcher, S.E.* Dynamic Motions of the HIV-1 Frameshift Site RNA. *Biophysical Journal* **2015**, 108(3), 644-654. DOI: [10.1016/j.bpj.2014.12.006](https://doi.org/10.1016/j.bpj.2014.12.006)
7. Low, J.T.; Garcia-Miranda, P.; **Mouzakis, K.D.**; Gorelick, R.J.; Butcher, S.E.; Weeks, K.M.* Structure and Dynamics of the HIV-1 Frameshift Element RNA. *ACS Biochemistry* **2014**, 53(26), 4282-4291. DOI: [10.1021/bi5004926](https://doi.org/10.1021/bi5004926)

Published prior to Dr. Mouzakis' independent faculty career:

8. **Mouzakis, K.D.**; Lang, A.L.; Vander Meulen; K.A., Easterday, P.D.; Butcher, S.E.* HIV-1 Frameshift Efficiency is Primarily Determined by the Stability of Base Pairs Positioned at the mRNA Entrance Channel of the Ribosome. *Nucleic Acids Research* **2013**, 41(3), 1901-1913. DOI: [10.1093/nar/gks1254](https://doi.org/10.1093/nar/gks1254)
9. Marcheschi, R.J.; **Mouzakis, K.D.**; Butcher, S.E.* Selection and Characterization of Small Molecules that Bind the HIV-1 Frameshift Site RNA. *ACS Chemical Biology* **2009**, 4(10): 844-854. DOI: [10.1021/cb900167m](https://doi.org/10.1021/cb900167m)

Book Chapters (published prior to Dr. Mouzakis' independent faculty career):

1. **Mouzakis, K.D.**; Burke, J.E.; Butcher, S.E. Investigating RNAs Involved in Translational Control by NMR and SAXS. In *Biophysical Approaches to Translational Control of Gene Expression*; J. D. Dinman (Ed.); Springer New York: London, England, 2013; pp 141-172. DOI: [10.1007/978-1-4614-3991-2_8](https://doi.org/10.1007/978-1-4614-3991-2_8)

External Grants:Awarded when Dr. Mouzakis held the rank of Associate Professor:

1. "Shape-based differentiation of RNA elements using small molecules", NSF Research Opportunity Award to Dr. Amanda Hargrove (PI) NSF CAREER Award (1750375) to support **Kathryn Mouzakis (Senior Personnel)**, 09/19/2022 – 08/01/2023, \$64,827.

Submitted when Dr. Mouzakis held the rank of Associate Professor and not funded:

- "From Tenure to Chair to Dean (and Beyond): Faculty Success and Leadership", 2025 Clare Boothe Luce Program for Women in STEM Proposal, Mahsa Ebrahim (Co-PI), Suzanne Larson (Co-PI), **Kathryn Mouzakis (Co-PI)**, Robin Wilson (Co-PI), and Delaram Yazdanehpas (Co-PI), application for \$490,000, submitted on 09/02/2025.
- "Never Have I Ever: Creating an Interinstitutional Peer Mentor and Faculty Support Network for Undergraduate Transfer Students in the Physical Sciences", RCSA Cottrell Scholar Collaborative (CSC) Proposal, **Kathryn Mouzakis (Co-PI)** and Carla Forlich (Co-PI), application for \$25,000, submitted on 08/31/2025.

Awarded when Dr. Mouzakis held the rank of Assistant Professor:

2. "Targeting the SARS-CoV-2 Frameshift Site Pseudoknot," **Kathryn Mouzakis (PI, team lead)**, Amanda Hargrove (PI), Research Corporation for Science Advancement (RCSA) COVID Initiative Award, 06/15/2020 – 06/14/2023, KDM and AEH each received \$55,000 awards from RCSA.
3. "Measuring HTLV-1 programmed ribosomal frameshifting *in vivo*," **Kathryn Mouzakis (Co-PI)**, Jeffrey Kieft (Co-PI), RCSA CSC Catalyzing Joint Research Award, 05/05/2019 – 05/04/2022, \$3,000.
4. "Seeding inclusive local networks for domain-specific research and teaching," **Kathryn Mouzakis (Co-PI)**, Mary Beth Anzovino (Co-PI), Aaron Romanowsky (Co-PI), RCSA CSC MSI/PWI Partnerships Award, 08/24/2018 – 08/23/2020, \$6,000.
5. "Structural Basis of -1 Programmed Ribosomal Frameshifting by the Human T-cell Lymphotropic Virus Type I RNA," **Kathryn Mouzakis (PI)**, RCSA Cottrell Scholar Award, 07/01/2017 – 12/31/2022, \$100,000.
6. "Structural Basis of -1 Programmed Ribosomal Frameshifting by the Human T-cell Lymphotropic Virus Type I RNA," **Kathryn Mouzakis (PI)**, NIH Support of Competitive Research (SCORE) Pilot Project Award (SC2), 02/01/2017 – 08/15/2018, \$300,000 awarded (\$225,978 used before the award was relinquished when the PI moved to LMU).

Internal Grants Awarded (Loyola Marymount University (LMU) & Fort Lewis College (FLC)):Awarded when Dr. Mouzakis held the rank of Associate Professor:

1. "Evaluating the correlation between an HTLV-1 RNA structure's stability and its function," LMU Seaver College Faculty Fellowship, **Kathryn Mouzakis (PI)**, Spring 2026, two-course release.

2. "Increasing access to undergraduate research experiences as a matter of justice", LMU Faith and Justice Research Grant, **Kathryn Mouzakis (PI)**, 06/01/2025 – 10/01/2025, \$4,000.
3. "Identifying RNA structural determinants of HTLV-1 -1 PRF," LMU Seaver College Continuing Research Grant, **Kathryn Mouzakis (PI)**, 06/01/2023 – 10/01/2023, \$5,000.

Awarded when Dr. Mouzakis held the rank of Assistant Professor:

4. "Travel support to attend the 2019 Cottrell Scholars Conference," LMU Center for Teaching Excellence Travel Grant, **Kathryn Mouzakis (PI)**, 06/01/2019 – 11/30/2019, \$500.
5. "Introduction to Virology," LMU Seaver College Course Development Grant, **Kathryn Mouzakis (PI)**, 07/01/2019 – 10/01/2019, \$5,000.
6. "Determining the Mechanism of Frameshifting in Human T-cell Leukemia Virus," FLC Faculty Development Grant for Traditional Research / Scholarship, **Kathryn Mouzakis (PI)**, 01/01/2014 – 12/31/2017, \$9,450.
7. "Integrating Undergraduate Research into the Curriculum," FLC Faculty Development Grant for Teaching Innovation, Pedagogy and Assessment, **Kathryn Mouzakis (PI)**, 10/01/2014 – 12/31/2017, \$4,400.
8. "Determining the Mechanism of Frameshifting in Human T-cell Leukemia Virus," FLC Foundation Grant, **Kathryn Mouzakis (PI)**, 01/01/2015 – 12/31/2016, \$4,223.
9. "Integrating Undergraduate Research into the Curriculum," FLC Foundation Grant, **Kathryn Mouzakis (PI)**, 01/01/2015 – 12/31/2016, \$3,500.
10. "Flipped Classrooms at FLC," FLC Teaching Empowerment Teams, **Kathryn Mouzakis (Co-PI)**, Kenny Miller (Co-PI), Carl Lienert (Co-PI), 09/01/2014 – 12/31/2014, \$500.
11. "Testing an Active Learning Environment versus Traditional Classroom Lectures in First Semester General Chemistry Courses," FLC Teaching Empowerment Teams, **Kathryn Mouzakis (Co-PI)**, Aimee Morris (Co-PI), 09/01/2013 – 12/31/2013, \$500.

Presentations (undergraduate mentees are underlined, presenting author(s)*):

Given when Dr. Mouzakis held the rank of Associate Professor:

1. Swanson Hay, E.G.; Maille, M.M.; Zafferani, M.; Puray-Chavez, M.; Xia, M.; Luu, T.B. Muralidharan, D.; Li, M.L.; Brewer, G.; Kutluay, S.B.; **Mouzakis, K.D.***; Hargrove, A.E. "SARS-CoV-2 Antivirals Identified from Small Molecule Modulators of Programmed –1 Ribosomal Frameshifting," (poster presentation). The 31st Annual Meeting of the RNA Society, Montreal, Canada, May 27, 2026.
2. **Mouzakis, K.D.*** "Programmed -1 Ribosomal Frameshifts Provide Alternate Paths to a Thriving Undergraduate Research Program," (invited oral presentation). The 43rd Steenbock Symposium: *Le Tour de RNA*, Madison, WI, USA, May 16, 2026.
3. **Mouzakis, K.D.*** "Inclusive Pedagogy in STEM", (oral presentation) Seaver College Inclusive Teaching Symposium, Los Angeles, CA, USA, May 6, 2026.
4. **Mouzakis, K.D.*** "SARS-CoV-2 Antivirals Identified from Small Molecule Modulators of Programmed –1 Ribosomal Frameshifting," (invited oral presentation). Pepperdine University Natural Science Seminar, Malibu, CA, USA, February 10, 2026.
5. **Mouzakis, K.D.*** "Celebrating 10 years of Scientific Discovery in the Life Sciences Building," (invited oral presentation). Seaver Spotlight, Los Angeles, CA, USA, October 8, 2025.
6. **Mouzakis, K.D.*** "Increasing Access to Undergraduate Research Experiences at Loyola Marymount University through an RNA-centric Course-based Undergraduate Research

- Experience in an Advanced Biochemistry Lab Course,” (oral presentation). The 30th Annual Meeting of the RNA Society, San Diego, CA, USA, May 29, 2025.
7. **Mouzakis, K.D.*** “Applying basic science research in pursuit of SARS-CoV-2 antivirals,” (oral presentation). Discover BMB 2024, The Annual Meeting of the American Society for Biochemistry and Molecular Biology (ASBMB), San Antonio, TX, USA, March 23, 2024.
 8. **Mouzakis, K.D.*** “AI in the Classroom: Practical Uses,” (invited oral presentation). LMU Teaching with Technology, Los Angeles, CA, USA, February 10, 2023.
 9. Weaver, K. *; Marino, B. *; **Mouzakis, K.D.***; Tarleton, H. * “Faculty Mindset Shift and Data Stewardship for Collective Accountability to PEER Student Experiences and Success,” (oral presentation). HHMI Driving Change Grant Awardee Meeting, Chevy Chase, MD, USA, November 11, 2022.
 10. **Mouzakis, K.D.*** “Applying basic science research in pursuit of SARS-CoV-2 antivirals,” (oral presentation). RNA@PUI Zoom Seminar Series, Online, October 20, 2022.
 11. **Mouzakis, K.D.*** “Applying basic science research in pursuit of SARS-CoV-2 antivirals,” (invited oral presentation). Santa Clara University Department of Chemistry and Biochemistry Seminar Series, Santa Clara, CA, USA, October 14, 2022.
 12. **Mouzakis, K.D.***, Soliman, M.E.; Robinson, L.K.; Maille, M.M.; Egekeze, C.C.; Salamon, R.J. “Revealing a Moderate Correlation Between the HTLV-1 *gag-pro* Frameshift Site Stem-loop Thermodynamic Stability and its Frameshift Efficiency,” (poster presentation). The 41st Annual Meeting of the American Society for Virology, Madison, WI, USA, July 18, 2022.
 13. Soliman, M.E.; Robinson, L.K.; Maille, M.M.; Egekeze, C.C.; Salamon, R.J.; **Mouzakis, K.D.*** “Revealing a Moderate Correlation Between the HTLV-1 *gag-pro* Frameshift Site Stem-loop Thermodynamic Stability and its Frameshift Efficiency,” (poster presentation). Bioorganic Gordon Research Conference, Andover, NH, USA, June 13, 2022.
 14. Soliman, M.E.; Robinson, L.K.; Maille, M.M.; Egekeze, C.C.; Salamon, R.J.; **Mouzakis, K.D.*** “Revealing a Moderate Correlation Between the HTLV-1 *gag-pro* Frameshift Site Stem-loop Thermodynamic Stability and its Frameshift Efficiency,” (poster presentation). The 27th Annual Meeting of the RNA Society, Boulder, CO, USA, June 2, 2022.
 15. **Mouzakis, K.D.*** “How a basic scientific research program focused on the fundamentals of viral RNA structure-stimulated translational reprogramming pivoted to applied science in response to the SARS-CoV-2 pandemic” (invited oral presentation). Locus Biosciences CRISPR Seminar Series, Durham, NC April 1, 2022.

Given **after** Dr. Mouzakis submitted her application for promotion to Associate Professor to the LMU Department of Chemistry and Biochemistry:

16. **Mouzakis, K.D.*** “An RNA Pseudoknot Stimulates HTLV-1 *pro-pol* Programmed -1 Ribosomal Frameshifting,” (invited oral presentation). The 2021 ASBMB Annual Meeting, Online, April 26, 2021.
17. **Mouzakis, K.D.*** “An RNA Pseudoknot Stimulates HTLV-1 *pro-pol* Programmed -1 Ribosomal Frameshifting,” (invited oral presentation). The Ohio State University Center for Retrovirus Research Seminar Series, Online, December 17, 2020.

Given when Dr. Mouzakis held the rank of Assistant Professor:

18. **Mouzakis, K.D.*** “An RNA Pseudoknot Stimulates HTLV-1 *pro-pol* Programmed -1 Ribosomal Frameshifting,” (oral presentation). Cold Spring Harbor Laboratory Virtual Conference: Retroviruses, Online, May 20, 2020.
19. **Mouzakis, K.D.*** “Active learning in SCSE spaces,” (invited oral presentation). LMU Center for Teaching Excellence New Faculty Seminar Series, Los Angeles, CA, USA, October 31, 2019.

20. **Mouzakis, K.D.*** "An RNA Pseudoknot Stimulates HTLV-1 *pro-pol* Programmed -1 Ribosomal Frameshifting," (invited oral presentation). The University of Southern California Chemical Biology Seminar Series, Los Angeles, CA, USA, October 10, 2019.
21. **Thulson, E.**; **Hartwick, E.W.**; **Cooper-Sansone, A.**; **Williams, M.A.C.**; **Kieft, J.S.**; **Mouzakis, K.D.*** "An RNA Pseudoknot Stimulates HTLV-1 *pro-pol* -1 Programmed Ribosomal Frameshifting," (poster presentation). Keystone Positive-Strand RNA Viruses Symposia, Killarney, Ireland, June 9-13, 2019.
22. **Mouzakis, K.D.*** "An RNA Pseudoknot Stimulates HTLV-1 *pro-pol* Programmed -1 Ribosomal Frameshifting," (invited oral presentation). Haverford Department of Chemistry Seminar Series, Haverford, PA, USA, April 18, 2019.
23. **Mouzakis, K.D.*** "Poll Everywhere Technology Integration Workshop," (invited oral presentation) LMU Center for Teaching Excellence Seminar Series, Los Angeles, CA, USA, February 22, 2019.
24. **Mouzakis, K.D.*** "Reprogramming the Ribosome: Programmed Ribosomal Frameshifting in HTLV-1," (invited oral presentation). The Harvey Mudd College Biology Colloquium, Claremont, CA, USA, September 26, 2018.
25. **Finke, E.A.**; **Chadeayne, D.P.**; **Durnford, K.M.**; **Hartwick, E.W.**; **Chapman, E.G.**; **Cooper-Sansone, A.**; **Williams, M.A.C.**; **Contreras, U.J.**; **Davis, S.S.**; **Greyeyes, S.D.**; **Knewitz, A.P.**; **Stelmaszek, J.A.**; **Atene, A.L.**; **Joe, N.S.**; **Dailey, L.A.**; **Broad, A.**; **Mackenzie, J.**; **Kieft, J.S.**; **Mouzakis, K.D.*** "An RNA Pseudoknot Stimulates Human T-cell Lymphotropic Virus Type 1 *pro-pol* -1 Programmed Ribosomal Frameshifting," (poster presentation). The 23rd Annual Meeting of the RNA Society, Berkeley, CA, USA, May 29 – June 2, 2018.
26. **Mouzakis, K.D.*** "Getting Involved in Undergraduate Research," (invited oral presentation). The FLC ASBMB Student Chapter: Discovering Research Series, Durango, CO, USA, March 13, 2018.
27. **Mouzakis, K.D.*** "Solving a Problem of Supply and Demand: Increasing Availability of Undergraduate Research Experiences (UREs) by Integrating Course-based UREs (CUREs) into the Biochemistry Curriculum," (oral presentation). Cottrell Scholar Conference: More Viewpoints, Better Science, Tucson, AZ, USA, July 12 – 14, 2017.
28. **Mouzakis, K.D.*** "Determination of the HTLV-1 *pro-pol* Frameshift Site Secondary Structure," (poster presentation). Cottrell Scholar Conference: More Viewpoints, Better Science, Tucson, AZ, USA, July 12 – 14, 2017.
29. **Finke, E.A.**; **Durnford, K.M.**; **Chapman, E.G.**; **Cooper-Sansone, A.**; **Dailey, L.A.**; **Contreras, U.J.**; **Broad, A.**; **Mackenzie, J.**; **Kieft, J.S.**; **Mouzakis, K.D.*** "Determination of the HTLV-1 *pro-pol* Frameshift Site Secondary Structure," (poster presentation). The 22nd Annual Meeting of the RNA Society, Prague, Czech Republic, May 30 – June 3, 2017.
30. **Finke, E.A.**; **Chadeayne, D.P.**; **Durnford, K.M.**; **Chapman, E.G.**; **Davis, S.S.**; **Greyeyes, S.D.**; **Knewitz, A.P.**; **Stelmaszek, J.A.**; **Broad, A.**; **Mackenzie, J.**; **Kieft, J.S.**; **Mouzakis, K.D.*** "Determination of the HTLV-1 *pro-pol* Frameshift Site Secondary Structure," (poster presentation). Gordon Research Conference on Post-Transcriptional Gene Regulation, Stowe, VT, USA, July 10 – 15, 2016.
31. **Mouzakis, K.D.*** "An RNA Research-based Advanced Biochemistry Laboratory Course: Design, Implementation, and Outcomes," (poster presentation). The 2015 Annual Meeting of the ASBMB, Boston, MA, USA, March 28 – April 1, 2015.

Given prior to Dr. Mouzakis independent faculty career:

32. **Mouzakis, K.D.***; **Cordes, M.***; **Wesley, M.***; **Keating, J.***; **Jose, G.***; **Berger, B.***; **Sibley, S.***; **Forest, K.T.** "A Response to the NSF Call to Action in Undergraduate Biology Education," (poster presentation). Teaching and Learning Symposium, Madison, WI, USA, May 22-23, 2013.

33. **Mouzakis, K.D.*** “The Role of RNA Structural Stability in HIV-1 -1 Programmed Ribosomal Frameshifting,” (invited oral presentation). Department Seminar, Carroll University, Waukesha, WI, USA, October 3, 2012.
34. **Mouzakis, K.D.*** “HIV-1 Frameshift Efficiency is Primarily Determined by the Stability of Three Base Pairs Positioned at the mRNA Entrance Channel of the Ribosome,” (oral presentation). The 17th Annual Meeting of the RNA Society, Ann Arbor, MI, USA, May 29 – June 2, 2012.
35. **Mouzakis, K.D.***; Marcheschi, R.J.; Tonelli, M.; Butcher, S.E. “Orientation and Dynamics of the HIV-1 Frameshift RNA Stem-Loop are Influenced by Counter-Ion and Ligand Interactions.” (Poster presentation). The 35th Steenbock Symposium: Advances in Biomolecular NMR, Madison, WI, USA, June 26 – 28, 2011.
36. **Mouzakis, K.D.***; Butcher, S.E. “The Role of the HIV-1 RNA Structure in Frameshifting Suggests Mechanical Stability Generally Regulates Frameshifting via Kinetic Control of Translation,” (poster presentation). Post-transcriptional Control: mRNA Translation, Localization, and Turnover, Edinburgh, UK, June 8 – 10, 2010.
37. **Mouzakis, K.D.***; Marcheschi, R.J.; Tonelli, M.; Butcher, S.E. “The Effects of Counter-ions and Ligands on the Orientation and Dynamics of the HIV-1 FS Stem-Loop,” (poster presentation). NIH Meeting of Groups Studying the Structures of AIDS-Related Systems, Bethesda, MD, USA, June 25 – 26, 2009.
38. **Mouzakis, K.D.***; Wu, T.; Haushalter, K.A. “Thermolability and Compromised Excision Activity of Polymorphic Forms of hOGG1,” (poster presentation). Keystone Symposium on Genome Instability and Repair, Breckenridge, CO, USA, January 17 – 21, 2007.

Student Presentations (undergraduate co-authors are underlined, presenting authors *):

Given when Dr. Mouzakis held the rank of Associate Professor:

1. Outley, N.D.*; Ibarra, G.E.; **Mouzakis, K.D.** “Quantifying the Impact of Stem-Loop Length on the HTLV-1 *gag-pro* Frameshift Efficiency,” (oral presentation). The 18th Annual LMU Undergraduate Research Symposium (URS), Los Angeles, CA, USA, March 20, 2026.
2. Rocha-Martin, N.A.*; Outley, N.D.; Ibarra, G.E.; **Mouzakis, K.D.** “Understanding the role of the HTLV-1 *gag-pro* frameshift site stem-loop thermodynamic stability in programmed –1 ribosomal frameshifting,” (poster presentation). The 18th Annual LMU URS, Los Angeles, CA, USA, March 20, 2026.
3. Gomez, M.S.*; Yamegni, M.E.; Covington, A.L.; **Mouzakis, K.D.** “Measuring frameshift efficiency of SARS-CoV-2 frameshift site using dual fluorescence reporter proteins,” (poster presentation). The 18th Annual LMU URS, Los Angeles, CA, USA, March 20, 2026.
4. Melton, K.A.*; **Mouzakis, K.D.** “Do base-triples form in the HTLV-1 *pro-pol* frameshift site?”, (poster presentation). The 18th Annual LMU URS, Los Angeles, CA, USA, March 20, 2026.
5. Chavez, A.R.*; Soedarmono, A.W.*; Bekele, L.B.; Rocha-Martin, N.A.; Maille, M.M.; **Mouzakis, K.D.** “Developing a protocol for evaluating *in vitro* transcribed RNA integrity using the Agilent 4150 TapeStation,” (poster presentation). The 18th Annual LMU URS, Los Angeles, CA, USA, March 20, 2026.
6. Chavez, A.R.*; Bekele, L.B.*; Rocha-Martin, N.A.; Maille, M.M.; **Mouzakis, K.D.** “Developing a protocol for evaluating *in vitro* transcribed RNA integrity using the Agilent 4150 TapeStation,” (poster presentation). The 2025 Southern California Conference for Undergraduate Research (SCCUR), Camarillo, CA, USA, November 22, 2025.
7. Rocha-Martin, N.A.*; Outley, N.D.; Ibarra, G.E.; **Mouzakis, K.D.** “Understanding the role of the HTLV-1 *gag-pro* frameshift site stem-loop thermodynamic stability in programmed

- 1 ribosomal frameshifting,” (poster presentation). The 2025 Annual Biomedical Research Conference for Minoritized Scientists (ABRCMS), San Antonio, TX, November 22, 2025.
8. Outley, N.D.*; Ibarra, G.E.; **Mouzakis, K.D.** “Quantifying the Impact of Stem-Loop Length on the HTLV-1 *gag-pro* Frameshift Efficiency,” (poster presentation). The 2025 ABRCMS, San Antonio, TX, November 22, 2025.
 9. Outley, N.D.*; Ibarra, G.E.; **Mouzakis, K.D.** “Quantifying the Impact of Stem-Loop Length on the HTLV-1 *gag-pro* Frameshift Efficiency,” (poster presentation). The 2025 Seaver Summer Research Poster Session, Los Angeles, CA, June 26, 2025.
 10. Yamegni, M.E.*; Covington, A.L.; Gomez, M.S.; **Mouzakis, K.D.** “Understanding the Role of the Stem-Loop Structure in HTLV-1 *gag-pro* frameshift efficiency,” (oral presentation). The 47th Annual West Coast Biological Sciences Undergraduate Research Conference (WCBSURC), San Marcos, CA, USA, April 12, 2025.
 - Received an Outstanding Oral Presentation Award.
 11. Gomez, M.S.*; Yamegni, M.E.; Covington, A.L.; **Mouzakis, K.D.** “Measuring frameshift efficiency of SARS-CoV-2 frameshift site using dual fluorescence reporter proteins”, (poster presentation). The 47th Annual WCBSURC, San Marcos, CA, USA, April 12, 2025.
 12. Melton, K.A.*; **Mouzakis, K.D.** “Do base-triples form in the HTLV-1 *pro-pol* frameshift site?”, (poster presentation). The 47th Annual WCBSURC, San Marcos, CA, USA, April 12, 2025.
 13. Yamegni, M.E.*; Covington, A.L.; Gomez, M.S.; **Mouzakis, K.D.** “Understanding the Role of the Stem-Loop Structure in HTLV-1 *gag-pro* frameshift efficiency,” (poster presentation). LMU 17th Annual URS, Los Angeles, CA, USA, March 21, 2025.
 14. Outley, N.D.*; Rocha-Martin, N.A.*; Gomez, M.S.*; Egekeze, C.C.; **Mouzakis, K.D.** “Quantifying the impact of stem-loop thermodynamic stability on the frameshift efficiency,” (poster presentation). LMU 17th Annual URS, Los Angeles, CA, USA, March 21, 2025.
 15. Melton, K.A.*; **Mouzakis, K.D.** “Do base-triples form in the HTLV-1 *pro-pol* frameshift site?”, (poster presentation). LMU 17th Annual URS, Los Angeles, CA, USA, March 21, 2025.
 16. Covington, A.L.*; Yamegni, M.E.*; Gomez, M.S.*; Maille, M.M.; **Mouzakis, K.D.** “Measuring the impact of an RNA stem-loop on the HTLV-1 *gag-pro* frameshift efficiency,” (poster presentation). National Conference on Undergraduate Research 2024, Long Beach, CA, USA, April 10, 2024.
 17. Melton, K.A.*; **Mouzakis, K.D.** “Do base-triples form in the HTLV-1 *pro-pol* frameshift site,” (poster presentation). LMU 16th Annual URS, Los Angeles, CA, USA, March 15, 2024.
 18. Covington, A.L.*; Yamegni, M.E.*; Gomez, M.S.*; Maille, M.M.; **Mouzakis, K.D.** “Measuring the impact of an RNA stem-loop on the HTLV-1 *gag-pro* frameshift efficiency,” (poster presentation). LMU 16th Annual URS, Los Angeles, CA, USA, March 15, 2024.
 19. Covington, A.L.*; Yamegni, M.E.; Gomez, M.S.; **Mouzakis, K.D.** “Measuring the impact of an RNA stem-loop thermodynamic stability on the HTLV-1 *gag-pro* frameshift efficiency,” (poster presentation). The 2023 Seaver Summer Research Poster Session, Los Angeles, CA, USA, June 22, 2023.
 20. Covington, A.L.*; Yamegni, M.E.; Gomez, M.S.; **Mouzakis, K.D.** “Measuring the impact of RNA stem-loop thermodynamic stability on the HTLV-1 *gag-pro* frameshift efficiency,” (oral presentation). RNA@PUI virtual Summer Student Seminar Series, Online, June 20, 2023.
 - Received an Outstanding Student Presentation Award.
 21. Maille, M.M.*; Zafferani, M.; Swanson, S.; Brewer, G.; Hargrove, A.E.; **Mouzakis, K.D.** “Investigating the Effect of Small Molecules on SARS-CoV-2 Programmed -1 Ribosomal Frameshifting,” (poster presentation). The 46th Annual WCBSURC, Los Angeles, CA, USA, April 22, 2023.

22. Covington, A.L.*; Maille, M.M.*; Yamegni, M.E.*; **Mouzakis, K.D.** “Measuring the impact of an RNA stem-loop on the HTLV-1 *gag-pro* frameshift efficiency,” (poster presentation). LMU 15th Annual URS, Los Angeles, CA, USA, March 17, 2023.
23. Soliman, M.E.*; Robinson, L.K.; Maille, M.M.; Egekeze, C.C.; Salamon, R.J.; **Mouzakis, K.D.** “Determining How Stem-loop Structure Thermodynamic Stability Influences Frameshift Efficiency at the HTLV-1 *gag-pro* Frameshift Site,” (poster presentation). HTLV 22 conference, Online, May 8-11, 2022.
24. Maille, M.M.*; **Mouzakis, K.D.** “Investigating the Role of Base-Triples in the HTLV-1 *pro-pol* Frameshift Site Pseudoknot,” (poster presentation). HTLV 22 conference, Online, May 8-11, 2022.
25. Soliman, M.E.*; Robinson, L.K.; Maille, M.M.; Egekeze, C.C.; Salamon, R.J.; **Mouzakis, K.D.** “Determining How Stem-loop Structure Thermodynamic Stability Influences Frameshift Efficiency at the HTLV-1 *gag-pro* Frameshift Site,” (poster presentation). The 45th Annual WCBSURC, San Diego, CA, USA, April 9, 2022.
26. Covington, A.L.*; **Mouzakis, K.D.** “Examining the Temperature-dependence of HTLV-1 *pro-pol* Programmed -1 Ribosomal Frameshifting,” (poster presentation). The 45th Annual WCBSURC, San Diego, CA, USA, April 9, 2022.
 - Received a Best Poster Presentation Award.
27. Maille, M.M.*; **Mouzakis, K.D.** “Investigating the Role of Base-Triples in the HTLV-1 *pro-pol* Frameshift Site Pseudoknot,” (poster presentation). The 45th Annual WCBSURC, San Diego, CA, USA, April 9, 2022.
28. Soliman, M.E.*; Robinson, L.K.; Maille, M.M.; Egekeze, C.C.; Salamon, R.J.; **Mouzakis, K.D.** “Determining How Stem-loop Structure Thermodynamic Stability Influences Frameshift Efficiency at the HTLV-1 *gag-pro* Frameshift Site,” (poster presentation). The 2022 Annual Meeting of the ASBMB, Philadelphia, PA, USA, April 2 – 5, 2022.
29. Maille, M.M.*; **Mouzakis, K.D.** “Investigating the Role of Base-Triples in the HTLV-1 *pro-pol* Frameshift Site Pseudoknot,” (poster presentation). The 2022 Annual Meeting of the ASBMB, Philadelphia, PA, USA, April 2 – 5, 2022.
30. Soliman, M.E.*; Robinson, L.K.; Maille, M.M.; Egekeze, C.C.; Salamon, R.J.; **Mouzakis, K.D.** “Determining How Stem-loop Structure Thermodynamic Stability Influences Frameshift Efficiency at the HTLV-1 *gag-pro* Frameshift Site,” (poster presentation). LMU 14th Annual URS, Los Angeles, CA, USA, March 19, 2022.
31. Maille, M.M.*; **Mouzakis, K.D.** “Investigating the Role of Base-Triples in the HTLV-1 Pseudoknot,” (poster presentation). LMU 14th Annual URS, Los Angeles, CA, USA, March 19, 2022.
32. Covington, A.L.*; **Mouzakis, K.D.** “Examining the Temperature-dependence of HTLV-1 *pro-pol* Programmed -1 Ribosomal Frameshifting,” (poster presentation). LMU 14th Annual URS, Los Angeles, CA, USA, March 19, 2022.
33. Melton, C.T.*; Hartwick, E.H.; Covington, A.L.; Kieft, J.S.; **Mouzakis, K.D.** “Evaluating the suitability of the pSGDLuc reporter plasmid to measure the HTLV-1 *pro-pol* frameshift efficiency in cells and *in vitro*,” (poster presentation). LMU 14th Annual URS, Los Angeles, CA, USA, March 19, 2022.
34. Soliman, M.E.*; Robinson, L.K.; Maille, M.M.; Egekeze, C.C.; Salamon, R.J.; **Mouzakis, K.D.** “Determining How Stem-loop Structure Thermodynamic Stability Influences Frameshift Efficiency at the HTLV-1 *gag-pro* Frameshift Site,” (oral presentation). The 2021 SCCUR, Online, November 20, 2021.
35. Maille, M.M.*; **Mouzakis, K.D.** “Investigating the Role of Base-Triples in the HTLV-1 Pseudoknot,” (poster presentation). The 2021 SCCUR, Online, November 20, 2021.
36. Covington, A.L.*; **Mouzakis, K.D.** “Examining the Temperature-dependence of HTLV-1 *pro-pol* Programmed -1 Ribosomal Frameshifting,” (poster presentation). The 2021 SCCUR, Online, November 20, 2021.

37. Melton, C.T.*; Hartwick, E.H.; Covington, A.L.; Kieft, J.S.; **Mouzakis, K.D.** "Evaluating the suitability of the pSGDLuc reporter plasmid to measure the HTLV-1 *pro-pol* frameshift efficiency in cells and *in vitro*," (poster presentation). The 2021 SCCUR, Online, November 20, 2021.
38. Stevens, K.A.*; Covington, A.L.; **Mouzakis, K.D.** "Measuring the Human OC43 Coronavirus Frameshift Efficiency," (poster presentation). The 2021 SACNAS National Diversity in STEM Digital Conference, Online, October 25-29, 2021.

Given after Dr. Mouzakis submitted her application for promotion to Associate Professor to the LMU Department of Chemistry and Biochemistry:

39. Stevens, K.A.*; Covington, A.L.; **Mouzakis, K.D.** "Measuring the Human OC43 Coronavirus Frameshift Efficiency," (poster presentation). The 2021 Virtual National McNair Conference at UCLA, Online, July 29, 2021.

Given when Dr. Mouzakis held the rank of Assistant Professor:

40. Egekeze, C.C.*; Soliman, M.E.; Robinson, L.K.; **Mouzakis, K.D.** "Investigating Spacer-length and Effects of RNA Stem-loop Thermodynamic Stability on HTLV-1 *gag-pro* -1 PRF Efficiency," (oral presentation). LMU 12th Annual URS, Online, April 18, 2020.
41. Soliman, M.E.*; Robinson, L.K.*; Egekeze, C.C.; **Mouzakis, K.D.** "Determining how Stem-loop Structure Thermodynamic Stability Influences Frameshift Efficiency at the HTLV-1 *gag-pro* Frameshift Site," (poster presentation). LMU 12th Annual URS, Online, April 18, 2020.
42. Robinson, L.K.*; Egekeze, C.C.*; Soliman, M.E.; **Mouzakis, K.D.** "Determining how stem-loop structure thermodynamic stability influences frameshift efficiency at the HTLV-1 *gag-pro* frameshift site," (poster presentation). The 2019 SCCUR, San Marcos, CA, USA, November 23, 2019.
43. Egekeze, C.C.*; Robinson, L.K.*; Soliman, M.E.*; **Mouzakis, K.D.** "Determination of the Relationship Between Thermodynamic RNA Stem-Loop Stability and Frameshift Efficiency," (poster presentation). LMU 11th Annual URS, Los Angeles, CA, USA, March 23, 2019.
44. Dailey, L.A.*; **Mouzakis, K.D.** "Assessing the Programmed -1 Ribosomal Frameshifting (-1 PRF) Efficiency of point mutations naturally occurring within the *pro-pol* frameshift site of HTLV-I," (invited oral presentation). FLC Undergraduate Research and Creative Activities Symposium (URCAS), Durango, CO, USA, April 19, 2018.
45. Cooper-Sansone, A.*; Williams, M.A.C.; Chadeayne, D.P.; Mylroie, E.A.; Contreras, U.J.; Davis, S.S.; Greyeyes, S.D.; Knewitz, A.P.; Stelmaszek, J.A.; Joe, N.S.; **Mouzakis, K.D.** "Evaluating the importance of pseudoknot formation to HTLV-1 *pro-pol* -1 programmed ribosomal frameshift stimulation," (poster presentation). The 2018 Annual Meeting of the ASBMB, San Diego, CA, USA, April 21 – 25, 2018.
46. Williams, M.A.C.*; Cooper-Sansone, A.; Chadeayne, D.P.; Mylroie, E.A.; Contreras, U.J.; Davis, S.S.; Greyeyes, S.D.; Knewitz, A.P.; Stelmaszek, J.A.; Joe, N.S.; **Mouzakis, K.D.** "Exploring the Significance of the Human T-cell Lymphotropic Virus Type-1 *pro-pol* Frameshift Site Pseudoknot," (poster presentation). The 2018 Annual Meeting of the ASBMB, San Diego, CA, USA, April 21 – 25, 2018.
47. White, E.*; Abrams, T.Y.; Banks, T.; Chadeayne, D.P.; Cooper-Sansone, A.; Contreras, U.J.; Dailey, L.A.; Davis, S.S.; Eades, A.E.; Greyeyes, S.D.; Harrison, J.A.; Hamilton, A.E.; Joe, N.S.; Knewitz, A.P.; Stelmaszek, J.A.; Mylroie, E.A.; Nash, H.P.; Williams, M.A.C.; **Mouzakis, K.D.** "Evaluation of *pro-pol* Frameshifting Efficiencies for Naturally Occurring Variants of HTLV-1," (poster presentation). The 2018 Annual Meeting of the ASBMB, San Diego, CA, USA, April 21 – 25, 2018.

48. Cooper-Sansone, A.*; Williams, M.A.C.; Chadeayne, D.P.; Mylroie, E.A.; Contreras, U.J.; Davis, S.S.; Greyeyes, S.D.; Knewitz, A.P.; Stelmaszek, J.A.; Joe, N.S.; **Mouzakis, K.D.** "Evaluating the importance of pseudoknot formation to HTLV-1 *pro-pol* -1 programmed ribosomal frameshift stimulation," (poster presentation). FLC URCAS, Durango, CO, USA, April 19, 2018.
49. Williams, M.A.C.*; Cooper-Sansone, A.; Chadeayne, D.P.; Mylroie, E.A.; Contreras, U.J.; Davis, S.S.; Greyeyes, S.D.; Knewitz, A.P.; Stelmaszek, J.A.; Joe, N.S.; **Mouzakis, K.D.** "Exploring the Significance of the Human T-cell Lymphotropic Virus Type-1 *pro-pol* Frameshift Site Pseudoknot," (poster presentation). FLC URCAS, Durango, CO, USA, April 19, 2018.
50. White, E.*; Abrams, T.Y.; Banks, T.; Chadeayne, D.P.; Cooper-Sansone, A.; Contreras, U.J.; Dailey, L.A.; Davis, S.S.; Eades, A.E.; Greyeyes, S.D.; Harrison, J.A.; Hamilton, A.E.; Joe, N.S.; Knewitz, A.P.; Stelmaszek, J.A.; Mylroie, E.A.; Nash, H.P.; Williams, M.A.C.; **Mouzakis, K.D.** "Evaluation of *pro-pol* Frameshifting Efficiencies for Naturally Occurring Variants of HTLV-1," (poster presentation). FLC URCAS, Durango, CO, USA, April 19, 2018.
51. Finke, E.A.*; **Mouzakis, K.D.** "Determination of the HTLV-1 *pro-pol* frameshift site RNA secondary structure," (poster presentation). American Society for Cell Biology, San Francisco, CA, USA, December 3 – 7, 2016.
52. Finke, E.A.*; **Mouzakis, K.D.** "Determination of the secondary RNA structure and its importance to the HTLV-1 *pro-pol* frameshift site," (poster presentation). FLC URCAS, Durango, CO, USA, April 21, 2016.
53. Salamon, R.J.*; Lee, J.A.; **Mouzakis, K.D.** "Minimal RNA Sequence Requirement of the HTLV-1 *gag-pro* RNA Frameshift Site," (poster presentation). FLC URCAS, Durango, CO, USA, April 21, 2016.
54. Caldera, H.I.*; Chadeayne, D.P.*; Eagle, F.W.*; Martin, E.*; Rios, J.R.*; Walker, M.A.*; **Mouzakis, K.D.** "Structure and Function of the HTLV-I *pro-pol* frameshift site," (poster presentation). FLC URCAS, Durango, CO, USA, April 21, 2016.
55. Absher, N.J.*; Block, C.W.*; Chavez, M.*; Joe, N.*; Lindquist-Kleissler, B.*; Mylroie, E.A.*; **Mouzakis, K.D.** "Studying the role of RNA secondary structure in Human T-cell Leukemia/Lymphotropic Virus Type 1 (HTLV-1) *gag-pro* -1 programmed frameshift site," (poster presentation). FLC URCAS, Durango, CO, USA, April 21, 2016.
56. Chadeayne, D.P.*; Davis, S.S.; Greyeyes, S.D.; Knewitz, A.P.; Stelmaszek, J.A.; **Mouzakis, K.D.** "Structure and Function of the HTLV-1 *pro-pol* Frameshift Site," (poster presentation). FLC URCAS, Durango, CO, USA, April 23, 2015.
57. Durnford, K.M.*; Chapman, E.G.; Atene, A.L.; Broad, A.; Mackenzie, J.; Yeager, D.L.; Kieft, J.S.; **Mouzakis, K.D.** "Determination of the HTLV-1 *pro-pol* Frameshift Site Secondary Structure," (poster presentation). FLC URCAS, Durango, CO, USA, April 23, 2015.
58. Chadeayne, D.P.*; Davis, S.S.; Greyeyes, S.D.; Knewitz, A.P.; Stelmaszek, J.A.; **Mouzakis, K.D.** "Structure and Function of the HTLV-1 *pro-pol* Frameshift Site," (poster presentation). The 2015 Annual Meeting of the ASBMB, Boston, MA, USA, March 28 – April 1, 2015.
59. Durnford, K.M.*; Chapman, E.G.; Atene, A.L.; Broad, A.; Mackenzie, J.; Yeager, D.L.; Kieft, J.S.; **Mouzakis, K.D.** "Determination of the HTLV-1 *pro-pol* Frameshift Site Secondary Structure," (poster presentation). The 2015 Annual Meeting of the ASBMB, Boston, MA, USA, March 28 – April 1, 2015.
60. Chadeayne, D.P.*; Davis, S.S.; Greyeyes, S.D.; Knewitz, A.P.; Stelmaszek, J.A.; **Mouzakis, K.D.** "Reprogramming the Ribosome: Structure and Function of the HTLV *pro-pol* Frameshift Site RNA," (poster presentation). FLC NSF FOCUSS Poster Session, Durango, CO, USA, June 23, 2014.

61. Lake, M.S.*; Lee, A.M.*; Mancha, S.R.*; **Mouzakis, K.D.** "The Effects of Local and Global RNA Stability on Frameshift Efficiency in the HTLV-II *gag-pro* Frameshift Site," (poster presentation). FLC URCAS, Durango, CO, USA, April 24, 2014.
62. Broad, A.J.*; McKenzie, J.T.*; **Mouzakis, K.D.** "Determining the secondary structure of the HTLV *pro-pol* frameshift site using SHAPE chemical probing," (invited oral presentation). FLC URCAS, Durango, CO, USA, April 24, 2014.
63. Davis, S.S.*; Greyeyes, S.D.*; Knewitz, A.P.*; Stelmaszek, J.A.*; **Mouzakis, K.D.** "Does Pseudoknot Formation play a role in HTLV-1 *pro-pol* Frameshift Efficiency?", (poster presentation). FLC URCAS, Durango, CO, USA, April 24, 2014.
64. Atene, A.L.*; Dayish, K.D.*; Warriner, M.M.*; Yeager, D.L.*; **Mouzakis, K.D.** "Determining the Secondary Structure of HTLV-1 *pro-pol* Frameshift Site using NMR and Native Gel Analysis," (poster presentation). FLC URCAS, Durango, CO, USA, April 24, 2014.

Undergraduate Research Students Mentored (co-author on a peer-reviewed publication*):

Loyola Marymount University

Mouzakis Lab Member:

1. Kamryn Higa (Biochemistry Major, 2029)	SP 2026 (12 wk. rotation) – present
2. Augustus Soedarmono (Science undeclared, 2029)	SP 2026
3. Michelle Rodas (Biochemistry Major, 2027)	SP 2026 (8 wk. rotation)
4. Saumaun Zadeh (Biology Major, 2028)	SP 2026 (8 wk. rotation)
5. Alexandar Chavez (B.S., Biochemistry, 2026)	FA 2025 – SP 2026
6. Neftali Rocha-Martin (B.S., Biology, 2026)	SP 2024 – SP 2026
7. Nigel Outley (B.S., Biochemistry, 2026)	SP 2024 – SP 2026
8. Marisa Gomez (B.S., Biology, 2026)	SU 2023 – SP 2026
9. Kennedy Melton (B.S., Biochemistry, 2026)	SP 2024 – FA 2025
10. Lucas Bekele (Biochemistry Major, 2027)	SU 2025 – FA 2025
11. Geraldray Ibarra (B.S., Biology, 2025)	SU 2025 – FA 2025
12. Mwanday Yamegni (B.S., Biochemistry, 2025)	SP 2023 – SP 2025
13. Audrey Covington (B.S., Biochemistry, 2024)	SP 2023 – SP 2024, SU 2021 – FA 2021
14. Jaynia Garcia (B.S., Biochemistry, 2022)	FA 2021
15. Cole Melton (B.S., Biology, 2022)	SU 2021 – FA 2021
16. Kristal Stevens (Biochemistry Major, 2023)	SU 2021

Students above joined the Mouzakis lab after Dr. Mouzakis submitted her application for promotion to Associate Professor to the LMU Department of Chemistry and Biochemistry

17. *Madison Maille (B.S., Biochemistry, 2023)	SP 2020 – SP 2023
18. *Mary Soliman (B.S., Biochemistry, 2022)	SP 2019 – SP 2022
19. *Leila Robinson (B.S., Biology, 2022)	FA 2019 – SP 2022
20. Michael Liu (B.S., Biology, 2022)	SP 2020 – SP 2021
21. Carolyn Egekeze (B.S., Biology, 2020)	SP 2019 – SP 2020
22. Douglas Kitchen (B.S., Biochemistry, 2019)	FA 2018 – SP 2019
23. Katerina Harrop (B.S., Biochemistry, 2019)	FA 2018 – SP 2019

Fort Lewis College

Mouzakis Lab Member:

1. Walter Potter (B.S., Biochemistry, 2019)	SP 2018
2. Harry Spencer (B.S., Biochemistry, 2019)	SP 2018
3. Emily White (B.S., Biochemistry, 2018)	FA 2017 – SP 2018
4. *Marcus Williams (B.S., Biochemistry, 2020)	FA 2017 – SP 2018
5. *Andrew Cooper-Sansone (B.S., Cellular and Molecular Biology, 2018)	SP 2017 – SP 2018

6. *Eliza Thulson (formerly Finke) (B.S., Cellular & Molecular Biology, 2017)	SU 2015 – SP 2017
7. Erin Gaffney (B.S., Chemistry – Biochemistry, 2017)	SP 2016 – SP 2017
8. Uriah Contreras (B.S., Biochemistry, 2019)	SP – FA 2017
9. Elena Mylroie (B.S., Chemistry – Biochemistry, 2017)	SP 2017
10. Rebecca Salamon (B.S., Cellular & Molecular Biology, 2018)	SU 2015 – SP 2016
11. Leandrew Dailey (B.S., Biochemistry, 2018)	FA 2016
12. Devon Chadeayne (B.S., Chemistry – Biochemistry, 2017)	SU 2014 – SU 2015
13. Melanie Walker (B.S., Chemistry – Biochemistry, 2016)	SP 2015
14. Kathryn Durnford (B.S., Cellular & Molecular Biology, 2015)	SU 2014 – SP 2015
15. Antonia Atene (B.S., Chemistry – Biochemistry, 2015)	FA 2014 – SP 2015
16. Natalie Joe (B.S., Chemistry – Biochemistry, Cellular & Molecular Biology, 2016)	FA 2015
17. Jamie Lee (NSF FOCUSS student, San Juan CC, 2015)	SU 2015
18. Austin Derksen (B.S., Cellular & Molecular Biology, 2016)	SP 2015
19. Grace Sheridan (B.S., Cellular & Molecular Biology, 2016)	SP 2015
20. Melvina Lake (B.S., Chemistry – Biochemistry, 2015)	SP 2014
21. Amanda Broad (B.S., Chemistry – Biochemistry, 2015)	SP 2014
22. Jason Mackenzie (B.S., Chemistry – Biochemistry, 2015)	SP 2014

Undergraduate Students Mentored in a CURE:

The courses below were taught when Dr. Mouzakis held the rank of Associate Professor

LMU CHEM 373 SP 2025 (taught in-person), 22 students	Damian Barrios, Lauren Crumb, Vivi Dallmer, Alice de Sa Costa Pereira, Madrid Ghanavat, Sophia Gonzalez, Jonathan Hei, Isabella May, Kennedy Melton, Benard Santana, Abydale Sotelo, Zoe Castanon, Alexander Chavez, Spencer Heaton, Ryan Iglesias, Kevin Lee, Larissa Negom, Mandy Nguyen, Tram Nguyen, Nigel Outley, Brooke Stagg, Jennifer Uchendu
LMU CHEM 373 SP 2024 (taught in-person), 15 students	Elizabeth Camberos, Niav Chapman, Alexa De Anda, Samantha del Pozo, Kyra DuBois, Emma Khacherian, Philippe LeGuellec, Sophie Menendez, Tia Nguyen, Nimrat Sran, Alianna Torres, Dara Varasteh, Brooke Welch, Mwanday Yamegni, Anthony Zaldana
LMU CHEM 373 SP 2023 (taught in-person), 19 students	Isa Ajpop, Alexa Batingana, Alyssa Michalek, Ashley Apostol, Audrey Covington, Brianna Gaspar, Christopher Ibrahim, Clara Goolsby, Hailey Ivanson, Havana Campo, Jade Hagihara, John Omiya, Joseph Hecht, Joshua Poura, Kelvin Akinyemi, Kevin Aclan, Monique Loza, Natnael Hagos, Philippe LeGuellec, Veda Taneng

The course below was taught **after** Dr. Mouzakis submitted her application for promotion to Associate Professor to the LMU Department of Chemistry and Biochemistry

LMU CHEM 373 SP 2021 (taught online), 28 students	Kayode Akinntire, Alex Barber, Savana Basconillo, Siri Breckenridge, Lianlen Joy Distor, Sara Esteves, Jaynia Garcia, Annie Heckman, Reed Hessel, Thomas Kelly, Kaitlyn Kennedy, Cat Machado, Quinton Markett, Finn Mercer, Kiana Nues, Sophia Papadopoulos, Natalie Perez, Ohara Riffel, Tess Rodriguez, Alyssa Rosales, Khaliah Sanders, Ariana Say, Mary Soliman, Sidney Starr, Katie Szymanski, Michelle Thaxton, Aaliyah Tyson, Ryan Vargas
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The courses below were taught when Dr. Mouzakis held the rank of Assistant Professor

FLC CHEM 496 FA 2017 (taught in-person), 8 students	Tara Abrams, Terrance Banks, Leandrew Dailey, Austin Eades, Jacob Harrison, Adam Hamilton, Hannah Nash, Emily White
FLC CHEM 411 SP 2016 (taught in-person), 12 students	Nathan Absher, Cooper Block, Hector Caldera, Devon Chadeayne, Marlyn Chavez, Forrest Eagle, Natalie Joe, Brent Lindquist-Kleissler, Evita Martin, Elena Mylroie, Jeovanna Rios, Melanie Walker
FLC CHEM 411 SP 2014 (taught in-person), 11 students	Antonia Atene, Summer Davis, Kolette Dayish, Allison Knewitz, Shawn Greyeyes, Melvina Lake, Audrianna Lee, Serena Mancha, Jordan Stelmaszek, Marcus Warriner, Daniel Yeager

Additional Mouzakis Lab Student Accomplishments (select alumni):

Alexander Chavez (LMU, '26): American Chemical Society (ACS) Inorganic Chemistry Award (2026); Currently applying to entry-level positions in industry with long-term plans to apply to Ph.D. programs in Biochemistry.

Kennedy Melton (LMU, '26): LMU Department of Chemistry and Biochemistry First-Year Chemistry Award (2023); LMU Honors Semester Research Fellowship (SP 2024); LMU Undergraduate Honors Thesis (FA 2025); American Institute of Chemists Best Overall Graduating Senior in Biochemistry (2026); Currently a Junior Formulation Scientist at Concept Labs.

Neftali Rocha-Martin (LMU, '26): LMU Academic Community of Excellence (ACE) Cohort member (2023-2026); HHMI Research Learning Community (RLC) Participant (FA 2023) and Undergraduate Research Student (SP 2024); Rains Undergraduate Research Fellow (Summer Undergraduate Research Program (SURP) SU 2025); LMU ACE Scholar (2026); Currently applying to entry-level positions in industry with plans to apply to Medical School.

Nigel Outley (LMU, '26): LMU ACE Cohort member (2023-2026); HHMI RLC Participant (FA 2023) and Undergraduate Research Student (SP 2024); University of Colorado Anschutz RNA Bioscience Initiative (RBI) Summer Internship Program Participant (SU 2024); Seaver College Summer Opportunities for Advanced Research (SOAR) Fellow (SU 2025); Currently applying to post-baccalaureate fellowship programs.

Marisa Gomez (LMU, '26): LMU ACE Cohort member (2023-2026); Rains Undergraduate Research Fellow (SURP SU 2023); HHMI RLC Participant (FA 2023) and Undergraduate Research Student (SP 2024); LMU ACE Marshall Saucedo Rising Scholar Award (2024); LMU Undergraduate Honors Thesis (FA 2025); LMU Raymunde McKay, R.S.H.M. Award (2026); LMU ACE Scholar (2026); Currently applying to Medical School.

Mwaday Yamegni (LMU, '25): Rains Undergraduate Research Fellow (SURP SU 2023); WCBSURC Outstanding Oral Presentation Award (2025); Stanford University Clinical Observation and Medical Transcription Program Fellow (2026); Currently applying to Medical School.

Audrey Covington (LMU, '24): Rains Undergraduate Research Fellow (SURP SU 2021); Harbor-UCLA/LMU Summer Research Program (SU 2022); WCBSURC Best Poster Presentation Award (2022); Seaver College SOAR Fellow (SU 2023); RNA@PUI Outstanding

Student Presentation Award (2023); ACS Division of Analytical Chemistry Undergraduate Award in Analytical Chemistry (2024); Currently a M.D. student at the University of California Irvine.

Madison Maille (LMU, '23): LMU Honors Summer Research Fellowship (SU 2021); Rains Undergraduate Research Fellow (SURP SU 2021); LMU Presidential Scholar; Honors Ambassadorial Grant (2022); Amgen Scholars Program Participant at Duke University (SU 2022); American Institute of Chemists Best Overall Graduating Senior in Biochemistry (2023); LMU Undergraduate Honors Thesis (SP 2023); Mouzakis Lab peer-reviewed publication co-author (2026); NIH Predoctoral Individual National Research Service Awardee (2026); Currently a Ph.D. student in the Department of Chemistry and Biochemistry at the University of California San Diego.

Kristal Stevens (LMU, '23): McNair Scholar; Summer Research Opportunities at Harvard (SROH) Participant (SU 2022); LMU Department of Chemistry and Biochemistry Michael P. Geis Memorial Research Award (2023); Currently a Ph.D. student in Chemical Biology at the Scripps Research Institute.

Mary Soliman (LMU, '22): Rains Undergraduate Research Fellow (SURP SU 2019); Mouzakis Lab peer-reviewed publication co-author (2020); Seaver College SOAR Fellow (SU 2021); NIH Undergraduate Scholarship Program Recipient (2021); LMU Department of Chemistry and Biochemistry Michael P. Geis Memorial Research Award (2022); Completed post-baccalaureate fellowships at the NIH National Cancer Institute (2023) and at the National Institute of Child Health and Human Development (2025); Currently applying to Medical School.

Leila Robinson (LMU, '22): LMU Honors Summer Research Fellowship (Summer 2019); Rains Undergraduate Research Fellow (UROP SP 2019, SURP SU 2019); Mouzakis Lab peer-reviewed publication co-author (2020); Department of Chemistry and Biochemistry Undergraduate Award in Organic Chemistry (2020); REU Participant at the Fred Hutchinson Cancer Research Center (SU 2021); LMU Undergraduate Honors Thesis (FA 2021); Honors Ambassadorial Grant (2022); Currently a M.D. student at the Pennsylvania State University College of Medicine.

Jaynia Garcia (LMU, '22): ACS Division of Analytical Chemistry Undergraduate Award in Analytical Chemistry (2022); Completed a NIH Intramural NIAID Research Opportunities (INRO) post-baccalaureate fellowship (2024); Currently a Ph.D. student in the Biomedical Sciences at the University of California San Diego.

Cole Melton (LMU, '22): LMU Honors Summer Research Fellowship (SU 2021); Rains Undergraduate Research Fellow (SURP SU 2021); LMU Presidential Scholar; LMU Undergraduate Honors Thesis (FA 2021); Earned a M.B.A. from the Creighton University Heider College of Business (2025); Currently a M.D. student at the Creighton University School of Medicine.

Carolyn Egekeze (LMU, '20): University of Colorado Anschutz RBI Summer Internship Program Participant (2019); LMU Undergraduate Honors Thesis (SP 2020); LMU Presidential Citation (2020); Completed a NIH post-baccalaureate IRTA fellowship at the National Institute of Child Health and Human Development; Guest Speaker for the LMU Chemistry Seminar course (SP 2026); Currently a M.D./Ph.D. student in the University of California Irvine Medical Scientist Training Program.

Marcus Williams (FLC, '20): NIH MARC U-STAR Undergraduate Research Fellow (2018); FLC Student Travel Grant (2018); Mouzakis Lab peer-reviewed publication co-author (2020); Completed a post-baccalaureate fellowship in Biomedical Engineering at the Johns Hopkins University School of Medicine (2022); Currently a M.D./Ph.D. student in the Joint Medical Scientist Training Program at the Universities of California San Francisco and Berkeley.

Andrew Cooper-Sansone (FLC, '18): University of Colorado Anschutz RBI Summer Internship Program Participant (2017); FLC Student Research, Scholarly and Creative Activities Award (2017); Mouzakis Lab peer-reviewed publication co-author (2020); Currently a content creator and freelance writer.

Rebecca Salamon (FLC, '18): NSF Four Corners Undergraduate STEM Success Undergraduate Research Student (2015); Amgen Scholars Program Participant at Washington University in Saint Louis (SU 2017); Earned a Ph.D. in Genetics from the University of Wisconsin – Madison; Completed a postdoctoral fellowship at the University of California San Diego; Currently a Senior Scientist I, Data Science and Biomarkers, at HAYA Therapeutics.

Leandrew Dailey (FLC, '18): NIH MARC U-STAR Undergraduate Research Fellow (2016); Completed a Postbaccalaureate Research Fellow at Albert Einstein College of Medicine (2019); Earned a Ph.D. in Chemistry and Chemical Biology from the University of California San Francisco (2025); Current position is unknown.

Eliza Thulson (formerly Finke; FLC, '17): FLC Undergraduate Honors Thesis (2017); FLC Student Research, Scholarly and Creative Activities Award (2015, 2016); Awarded an NSF Graduate Research Fellowship (2019); Mouzakis Lab peer-reviewed publication co-author (2020); Earned a Ph.D. in Genetics and Molecular Biology from the University of North Carolina (2023); Currently a senior medical writer at Dexcom.

Erin Gaffney (FLC, '17): FLC Student Research, Scholarly and Creative Activities Award (2016); Earned a Ph.D. in Chemistry from the University of Utah (2021); Completed a postdoctoral fellowship at the University of Utah (2022); Currently working as a Life Sciences Innovation & IP Strategist at Shape Therapeutics Inc.

Natalie Joe (FLC, '16): NIH MARC U-STAR Undergraduate Research Fellow (2014); Earned a Ph.D. in Cellular and Molecular Medicine from the Johns Hopkins University School of Medicine (2022); Completed a postdoctoral fellowship at the National Cancer Institute (2026); Currently an Assistant Professor of Oncology at Roswell Park Comprehensive Cancer Center.

Kathryn Durnford (FLC, '15): FLC Student Research, Scholarly and Creative Activities Award (2014); Earned a M.D. and a M.B.A. from the University of Utah (2022); Currently a practicing physician at Billings Clinic.

Amanda Broad (FLC, '14): FLC Student Research, Scholarly and Creative Activities Award (2014); Earned a Ph.D. in Biochemistry and Molecular Biology from Colorado State University (2020); Currently a Senior Scientist at Arpeggio Biosciences.

SERVICE

LMU – University Service:

Strategic Plan Steering Committee – Committee Member
Sabbatical Review Committee – Interim Chair

05/2026 – present
SP 2023, SP 2026

Sabbatical Review Committee –	FA 2023 – SP 2024, SP 2025 – SP 2026
Committee Member	FA 2020 – SP 2021
LMU URS – Moderator	03/17/2023, 04/20/2026
Presidential Inauguration Committee – Academic Programming	FA 2025
Subcommittee Chair	
LMU Proposal Writing Academy – Grant Review Panelist	08/23/2023
Center for Teaching Excellence Event: Faculty Support for	02/08/2023
Mentoring Student Research – Panel Member	
Honors Advisory Council – Interim Committee Member	FA 2020 – SP 2021
Board of Trustees Committee on Academic Affairs –	12/06/2020
Guest Faculty Committee Panel Member	
Sabbatical Review Committee – Interim Committee Member	SP 2020

LMU – Seaver College of Science and Engineering (SCSE) Service:

College Curriculum Committee – Committee Member	SP 2024, SP 2025 – present
	FA 2019 – FA 2021, SP 2023
Become a LMU Lion: Seaver Dean’s Session Faculty	03/28/2026
Panelist	
Seaver DEIC National Academies Workshop –	02/12/2026
Introduced the Guest Speaker	
Seaver Spotlight Presenter	10/08/2025
HHMI Driving Change Core Team Member	11/2022 – 04/2024
College Curriculum Committee – Committee Co-Chair	AY 2023 – 2024
WCBSURC Moderator and Judge	04/22/2023
SCSE Family Day Speaker	02/17/2023
Experience LMU: Connect with Dean Choe, SCSE	10/19/2021
Faculty Panelist	
Guest Presentation to the Seaver Executive Advisory Board	01/14/2021

LMU – Department of Chemistry and Biochemistry Service:

Biochemistry Lecture (CHEM 370) 4-unit curriculum change coordinator	SU 2026 – present
First Year Advisor to Chemistry and Biochemistry Majors	SU 2026 – present
Ad Hoc Working Group on revision of the Departmental Merit Standards – Member	SU 2026 – present
Ad Hoc Working Group on Policy for Hiring Student Research Assistant(s) – Member	SP 2026 – present
Voluntary Mentor to a new Assistant Professor in Biochemistry	FA 2025 – present
Chemistry Instructor Search Committee – Committee Member	SP 2026
Peer teaching observer	FA 2021, SP 2026
CHEM 190 Presenter	Annually (since 2018)
Lead-instructor for Biochemistry Lab (CHEM 371)	FA 2021, FA 2023, FA 2025
Ad Hoc Working Group on Student Writing - Chair	SP 2025
Contributor to the Departmental ACS Periodic Report	SP 2025
Contributor to the Departmental 4-unit Curriculum Proposal	SP 2025
Assistant Professor of Biochemistry Search Committee –	01/2025
Committee Member	
Assistant Professor of Biochemistry Search Committee –	AY 2023 – 2024
Committee Chair	

Contributor to the revision of the 2023 Departmental Rank and Tenure Standards	SP 2023
Chemistry Instructor Search Committee – Committee Member	FA 2021
Department Assessment Committee – Committee Member	FA 2021
Department Assessment Committee – Interim Committee Member	FA 2019 – SP 2021
Department Ad Hoc Committee on Workload – Committee Member	FA 2018
Open House and Preview Day – Faculty Participant	FA 2018

FLC – University Service:

Faculty Senate Science Representative	AY 2016 – 2018
Safe Zone Ally	AY 2015 – 2018
FLC Business Competition – Advisory Board Member	AY 2015 – 2017
New Student Orientation Professor's Perspective – Panel Member	FA 2016

FLC – School of Arts and Sciences Service:

Faculty Mentor to two out-of-department Assistant Professors	AY 2016 – 2018
Undergraduate Scholarly and Creative Activities – Grant Reviewer	2015 – 2018
Assistant Professor in Mathematics Search – Committee Member	FA 2016
URCAS – Faculty Liaison	AY 2015 – 2016
Women in Science Student Organization – Faculty Advisor	FA 2015
Assistant Professor in Molecular Microbiology Search – Committee Member	FA 2014

FLC – Department of Chemistry and Biochemistry Service:

Department Seminar Series – Organizer	AY 2014 – 2018
Major Contributor to the Department Assessment Plan	AY 2013 – 2018
Chemistry Club – Faculty Advisor	AY 2015 – 2016
Assistant Professor in Physical Chemistry Search – Committee Member	FA 2015
Visiting Chemistry Instructor Search – Committee Member	SP 2014

Professional Service:

RCSA STAR/Impact Award – Review Panel Member	02/2026 – 03/2026
RCSA SEED and Cottrell Scholar Awards – Ad Hoc Reviewer	04/2025, 09/2025, 05/2026
RNA Society Mentor-Mentee Luncheon – Mentor	09/2018, 09/2019, 09/2021
	05/2025, 05/2026
	06/2017, 06/2018
Tenure and Promotion – External Reviewer	10/2021, 10/2022 (2 let.), 11/2025
Fulbright-Cottrell Award – Ad Hoc Reviewer	03/2024
CSC Fulbright-Cottrell Building Bridges Workshop – Facilitator	06/2023
CSC Workshop on Alternative Grading – Co-organizer	AY 2021 – 2022
Bioorganic Gordon Research Conference Nucleic Acid Technologies – Discussion Lead	06/15/2022
NSF Chemistry and Biology Division – Grant Reviewer	12/2021 – 02/2022
Chem Bio Connections Virtual Seminar Series – Co-organizer	05/2021 – 08/2021
ACS New Faculty Workshop – Facilitator	06/2021
Rustbelt RNA Meeting 2020: Solutions for Promoting DEI in Science – Panel Member	10/24/2020
Cottrell Scholars Regional Meeting – Committee Member	03/2020 – 07/2020
RCSA Board Meeting – Cottrell Scholars Panel Member	02/2019
CSC MSI/PWI Partnerships – Workshop Facilitator	07/2018

Gordon Research Seminar on Post-Transcriptional Gene Regulation – Career Panel Member	07/2016
National Academies Mountain West Summer Institute – Facilitator	06/2015
ASBMB Undergraduate Poster Competition – Judge	03/29/2015

HONORS AND AWARDS

RNA Society Research Fellowship	2022
LMU Honors Program Honorable Mentor Award	2022
ASV Teacher of Undergraduate Students Registration Award	2021, 2022
LMU Ascending Scholar Award	2021
RNA Society Travel Fellowship	2017, 2018
RCSA Cottrell Scholar Award	2017
Gordon Conference Travel Award	2016
FLC New Faculty Teaching Award	2016
ASBMB Faculty Travel Award	2015
National Academies Education Fellow in the Life Sciences	2013 – 2014
University of Wisconsin – Madison Honored Instructor Award	2012
HHMI Teaching Fellow	2011 – 2012
University of Wisconsin – Madison Biochemistry Travel Award	2010, 2012
University of Wisconsin – Madison Vilas Travel Grant	2010

PROFESSIONAL ACTIVITIES

LMU Center for Faculty Development: Leadership Workshop Series – Completed as a Leadership Cohort Member	AY 2025 – 2026
LMU HHMI Faculty Learning Community – Participant	AY 2025 – 2026
LMU HHMI STEM Equity Summer Institute – Participant	07/2025 – 08/2025
RNA Society – Member	2012, 2017 – 2020, 2022, 2025, 2026
ASBMB – Member	2015, 2022, 2024
LMU Proposal Writing Academy – Participant	08/2022
American Society for Virology – Member	2022
Biophysical Society – Member	2021
CSC Academic Leadership Training Workshop – Participant	02/2019
Council on Undergraduate Research (CUR) – Member	2015
CUR Proposal Writing Institute – Participant	07/2015
National Academies Mountain West Summer Institute – Participant	08/2013
Biochemical Society – Member	2010